

## CYCV15-CHANGYU pTOPO-TA Simple Kit

| Contents                       | 20 rxns<br>(CYCV1501) | 80 rxns<br>(CYCV1502) |
|--------------------------------|-----------------------|-----------------------|
| pTOPO-T Simple Vector(30ng/μl) | 20 μl                 | 80 μl                 |
| 1000bp Control (30ng/μl)       | 5 μl                  | 5 μl                  |
| 10 × Enhancer                  | 20 μl                 | 80 μl                 |

Note: All reagents, when stored at -20 °C, are stable for 12 months.

### ❖ Description

The Zero Background TA Simple Topoisomerase Cloning Kit is designed for fast cloning of DNA fragments up to 10 kb generated by *Taq* DNA polymerase which introduces single 3'-A overhang to the end of the DNA product. DNA fragments generated by restriction digestion or obtained by mechanical shearing can also be effectively cloned after end polishing using *Taq* DNA polymerase. It utilizes DNA strand transfer activity of *Vaccinia* virus topoisomerase I. *Vaccinia* virus DNA topoisomerase I forms a 3'-phosphoryl intermediate with the plasmid vector containing cleavage recognition motif of 5'CCCTT↓. Covalently bound topoisomerase I then transfers the incised vector DNA strand to the DNA fragment to be cloned with free 5'-OH termini. This transferring reaction is rapid and reproducible. The cloning vector pTOPO-TA Simple included in this kit is a high copy number plasmids engineered to tolerate mildly toxic genes. Regions flanking the cloning site of pTOPO-TA Simple vectors do not contain any common restriction sites which eliminates possible redundancy of restriction sites.

### ❖ Procedure

1. Set up the topoisomerase cloning reaction by mixing the reagents in the order shown.

|                                    |         |
|------------------------------------|---------|
| DNA Fragment or 1μl 1000bp control | 0.5-8μl |
| pTOPO-T Simple Vector              | 1μl     |
| 10 × Enhancer                      | 1μl     |
| ddH <sub>2</sub> O                 | Xμl     |
| Final Volume                       | 10μl    |

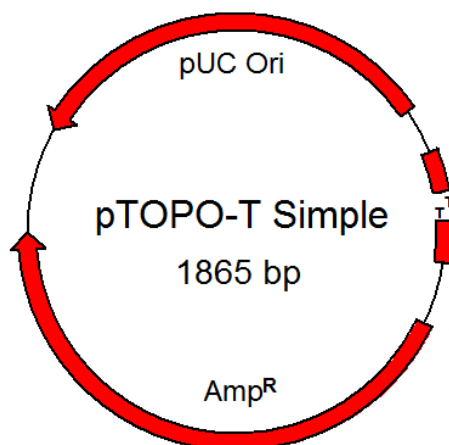
DNA fragments MUST NOT be 5'-phosphorylated. TA-ended DNA fragment is preferred. Highest cloning efficiency was observed from PCR fragments generated by *Taq* DNA polymerase or fragments tailored by *Taq* polymerase. 10-150 ng of

DNA fragments ranging from 100 bp to 5000 bp have been tested to give satisfactory results. Excessive amounts of DNA inserts e.g. >300ng, will significantly reduce cloning efficiency. Refer to the following table.

| Fragment size (bp) | Optimal amount (ng) |
|--------------------|---------------------|
| 100-1000           | 10-40               |
| 1000-2000          | 40-80               |
| 2000-5000          | 80-150              |

2. Mix the reaction gently and incubate for 5 minutes at room temperature between 15-30°C. Recent R&D data indicate incubation at this step produces no detectable benefits. In any case, do not let the incubation go beyond 5 minutes. Extended incubation for larger inserts up to 5 kb is unnecessary and may introduce background.
3. Add 5 µl of the cloning reaction into 50 µl chemically competent E. coli and mix gently. Do not mix by pipetting up and down.
4. Incubate on ice for 2-30 minutes.
5. Follow the competent cell manufacturer's instructions to complete the transformation.

❖ **Map of pTOPO-T Simple:**



❖ **Sequencing Primer:**

M13F: TGTAACGACGGCCAGT

M13R: CAGGAAACAGCTATGACC

❖ **Cloning site information:**

